# LZI Fentanyl II (Semi-Quantitative) Enzyme Immunoassay

 $\begin{array}{c} \textbf{REF} \\ 0580 \; (100/37.5 \; mL \; R_1/R_2 \; Kit) \\ 0581 \; (1000/375 \; mL \; R_1/R_2 \; Kit) \end{array}$ 





## For Forensic Use Only

## Lin-Zhi International, Inc.

#### **Intended Use**

The LZI Fentanyl II (Semi-Quantitative) Enzyme Immunoassay is intended for the semi-quantitative determination of norfentanyl in human urine at the cutoff value of 5 ng/mL when calibrated against norfentanyl. The assay is designed for use with a number of automated clinical chemistry analyzers. This is a non-FDA approved assay for Forensic Use Only and as such should not be repackaged for *in vitro* diagnostic use.

The assay provides only a preliminary analytical result. A more specific alternative chemical method (e.g., gas or liquid chromatography and mass spectrometry) must be used in order to obtain a confirmed analytical result. (1, 2). Clinical consideration and professional judgment should be exercised with any drug of abuse test result, particularly when the preliminary test result is positive.

## **Summary and Explanation of Test**

Fentanyl is an important opioid analgesic used widely in surgical operations and is a controlled substance (3). Fentanyl is most commonly encountered in the form of patches applied to the skin, as "lollipops" which can be dissolved in the mouth through the mucous membrane, or can be administered intravenously. It is 50-100 times stronger than morphine (4, 5) and cases of fentanyl abuse via intravenous injection, inhalation, oral, or nasal applications have been previously reported (6). Fentanyl is used in the treatment of acute and chronic pain, usually in patients who no longer respond to high doses of less potent opioids such as morphine or oxycodone. Due to its potency and wide availability as a prescribed drug, fentanyl has been abused and misused by health professionals, pain management patients, and recreational abusers (7).

Due to its short elimination half-life and approximately 90 % metabolism, fentanyl is difficult to detect in urine (8). Fentanyl undergoes extensive hepatic biotransformation to metabolites coming from hydrolysis, N-dealkylation, or hydroxylation reactions (9). In an intravenous dose of fentanyl, up to 85 % is excreted in urine over a three- to four- day period with 0.4-6 % eliminated as unchanged fentanyl and 26-55 % eliminated as the norfentanyl metabolite (10).

Fentanyl analogs also have high potency analgesic activities. Numerous reports have been published with modified fentanyl-related compounds abused as designer drugs (11-13).

Other recently available fentanyl analogs associated with abuse and severe intoxication include butyryl fentanyl and 4-fluorobutyryl fentanyl (14-18).

## **Assay Principle**

The LZI Fentanyl II (Semi-Quantitative) Enzyme Immunoassay is a homogeneous enzyme immunoassay ready-to-use liquid reagent. The assay is based on competition between drug in the sample and drug labeled with the enzyme glucose-6-phosphate dehydrogenase (G6PDH) for a fixed amount of antibody in the reagent (19). Enzyme activity decreases upon binding to the antibody, and the drug concentration in the sample is measured in terms of enzyme activity. In the absence of drug in the sample, fentanyl-labeled G6PDH conjugate is bound to antibody, and the enzyme activity is inhibited. On the other hand, when drug is present in the sample, antibody would bind to free drug; the unbound fentanyl-labeled G6PDH then exhibits its maximal enzyme activity. Active enzyme converts nicotinamide adenine dinucleotide (NAD) to NADH, resulting in an absorbance change that can be measured spectrophotometrically at 340 nm.

## **Reagents Provided**

Antibody/Substrate Reagent ( $R_1$ ): Contains a mouse monoclonal anti-fentanyl antibody, glucose-6-phosphate (G6P), nicotinamide adenine dinucleotide (NAD), stabilizers, and sodium azide (0.09 %) as a preservative. Enzyme-drug Conjugate Reagent ( $R_2$ ): Contains glucose-6-phosphate dehydrogenase (G6PDH) labeled with fentanyl in buffer with sodium azide (0.09 %) as a preservative.

#### Calibrators and Controls\*

\*Calibrators and Controls are sold separately and contain negative human urine with sodium azide as a preservative.

NORFENTANYL (Semi-Quantitative) Calibrators	REF
Negative Calibrator	0001
Low Calibrator: Contains 2.5 ng/mL norfentanyl	0552
Cutoff Calibrator: Contains 5 ng/mL norfentanyl	0553
Intermediate Calibrator: Contains 10 ng/mL norfentanyl	0554
High Calibrator: Contains 20 ng/mL norfentanyl	0555

NORFENTANYL (Semi-Quantitative) Controls	REF
Level 1 Control: Contains 3.75 ng/mL norfentanyl	0557
Level 2 Control: Contains 6.25 ng/mL norfentanyl	0558

## **Precautions and Warning**

- This test is for Forensic Use Only. This test should not be re-packaged for in vitro diagnostic use.
- · Harmful if swallowed.
- Reagent contains sodium azide as a preservative, which may form
  explosive compounds in metal drain lines. When disposing such reagents or
  wastes, always flush with a large volume of water to prevent azide buildup. See National Institute for Occupational Safety and Health Bulletin:
  Explosive Azide Hazards (20).
- Do not use the reagents beyond their expiration dates.

## **Reagent Preparation and Storage**

The reagents are ready-to-use. No reagent preparation is required. All assay components should be refrigerated at 2-8°C when not in use.

## **Specimen Collection and Handling**

Use fresh urine specimens for the test. If the sample cannot be analyzed immediately, it may be refrigerated at 2-8°C for up to four weeks (21) or at room temperature for up to four weeks (21, 22). For longer storage, keep sample frozen at -20°C and then thaw before use. Studies have shown norfentanyl samples in urine are stable at -20°C for up to six months (23). Sample stability claims were established by experimental data by the manufacturer or based on reference literature and only for the temperatures/time frames as stated in the method sheet. It is the responsibility of the individual laboratory to use all available references and/or its own studies to determine specific stability criteria for its laboratory.

Samples should be equilibrated to room temperature (18-25°C) for testing. Samples with high turbidity should be centrifuged before analysis. Adulteration may cause erroneous results. If sample adulteration is suspected, obtain a new sample and both samples should be forwarded to a laboratory for testing.

Handle all urine specimens as if they are potentially infectious.

## Instrument

Clinical chemistry analyzers capable of maintaining a constant temperature, pipetting sample, mixing reagents, measuring enzyme rates at 340 nm and timing the reaction accurately can be used to perform this homogeneous immunoassay.

Performance characteristics presented in this package insert have been validated on the Beckman Coulter AU480.

## Assay Procedure

Typical assay parameters used for the Beckman Coulter AU480 analyzer include a 20  $\mu L$  sample, 120  $\mu L$  of antibody reagent (R1), 45  $\mu L$  of enzyme conjugate reagent (R2), 12-16 reading frame, FIXED method, and 340 nm primary wavelength. If additional washing steps are required, reference analyzer specific parameter sheet.

For semi-quantitative analysis, use all five calibrators.

Recalibration should be performed after reagent bottle change or a change in calibrators or reagent lot. Two levels of controls are also available for monitoring the cutoff level: 3.75 ng/mL and 6.25 ng/mL.

## **Calibration and Quality Control**

Good laboratory practices recommend the use of at least two levels of control specimens (one positive and one negative control near the cutoff) to ensure proper assay performance. Controls should be run with each new calibration and after specific maintenance or troubleshooting procedures as detailed in the instrument system manual. Each laboratory should establish its own control frequency. If any trends or sudden change in control value are observed, review all operating parameters, or contact LZI technical support for further assistance. Laboratories should comply with all federal, state, and local laws, as well as all guidelines and regulations.

#### Results

**Note:** A preliminary positive test result does not necessarily mean a person took a specific drug and a negative test result does not necessarily mean a person did not take a specific drug. There are a number of factors that influence the reliability of drug tests.

Semi-Quantitative: The semi-quantitative mode is for purposes of (1) enabling laboratories to determine an appropriate dilution of the specimen for verification by a confirmatory method such as GC/MS, LC/MS or (2) permitting laboratories to establish quality control procedures. When an approximation of concentration is required, a calibration curve can be established with five calibrators. The concentration of norfentanyl in the sample may then be estimated from the calibration curve.

#### Limitations

- 1. Boric Acid at 1% w/v may cause false negative results. Boric Acid is not recommended as a preservative for urine.
- 2. Dextromethorphan may cause false positive results at concentrations greater than 40,000 ng/mL.
- 3. A preliminary positive result from this assay indicates only the presence of norfentanyl and does not necessarily correlate with the extent of physiological and psychological effects (e.g., intoxication). This test is not intended for quantifying the individual analytes in samples.
- 4. A negative result does not necessarily mean a person did not abuse drugs.
- Care should be taken when reporting results, as numerous factors (e.g., fluid intake, endogenous or exogenous interferents) may influence the urine test result.
- Preliminary positive results should be confirmed by other affirmative, analytical methods (e.g., chromatography), preferably GC/MS or LC/MS.
- 7. The test is designed for use with human urine only.
- 8. The test is not for therapeutic drug monitoring.

#### **Typical Performance Characteristics**

The results shown below were performed with a single Beckman Coulter AU480 automated chemistry analyzer.

#### Precision:

<u>Qualitative analysis</u>: The following concentrations were evaluated. Typical qualitative results (measured by  $\Delta OD$ , mAU) are as follows:

Concentration	Within Run (N=22)			Total Precision (N=88)		
Concentration	Mean	SD	% CV	Mean	SD	% CV
0 ng/mL	0.0	0.0	N/A	0.0	0.2	N/A
1.25 ng/mL	15.5	3.0	20.2%	15.5	4.1	26.6%
2.5 ng/mL	35.1	2.9	8.4%	35.1	3.5	10.0%
3.75 ng/mL	58.8	3.0	5.1%	58.8	3.8	6.5%
5 ng/mL	81.0	3.3	4.1%	81.0	3.7	4.6%
6.25 ng/mL	105.5	2.7	2.5%	105.5	3.9	3.7%
7.5 ng/mL	128.0	2.9	2.3%	128.0	3.5	2.8%
8.75 ng/mL	150.1	3.2	2.2%	150.1	4.2	2.8%
10 ng/mL	171.3	4.3	2.5%	171.3	5.2	3.0%

5 ng/mL Cutoff		Within Ru	m (N = 22)	<b>Run-to-Run</b> (N = 88)	
Concentrati	% of	#	EIA	# Samples	EIA
on	Cutoff	Samples	Result	# Samples	Result
0 ng/mL	0 %	22	22 Neg	88	88 Neg
1.25 ng/mL	25 %	22	22 Neg	88	88 Neg
2.5 ng/mL	50 %	22	22 Neg	88	88 Neg
3.75 ng/mL	75 %	22	22 Neg	88	88 Neg
5 ng/mL	100 %	22	13 Neg/ 9 Pos	88	59 Neg/ 29 Pos
6.25 ng/mL	125 %	22	22 Pos	88	88 Pos
7.5 ng/mL	150 %	22	22 Pos	88	88 Pos
8.75 ng/mL	175 %	22	22 Pos	88	88 Pos
10 ng/mL	200 %	22	22 Pos	88	88 Pos

<u>Semi-quantitative analysis</u>: The following concentrations were determined with reference curves from 5 calibrators. Typical results were measured in ng/mL.

C	Within Run (N=22)			Total Precision (N=88)		
Concentration	Mean	SD	% CV	Mean	SD	% CV
0 ng/mL	0.2	0.1	N/A	0.2	0.1	N/A
1.25 ng/mL	1.4	0.1	10.5%	1.4	0.2	12.6%
2.5 ng/mL	2.5	0.1	4.6%	2.5	0.2	6.5%
3.75 ng/mL	3.6	0.1	3.7%	3.6	0.2	5.0%
5 ng/mL	4.8	0.1	2.9%	4.8	0.2	3.9%
6.25 ng/mL	6.1	0.2	3.0%	6.1	0.2	3.4%
7.5 ng/mL	7.4	0.2	2.5%	7.4	0.2	3.3%
8.75 ng/mL	8.6	0.2	1.8%	8.6	0.2	2.6%
10 ng/mL	9.9	0.2	2.3%	9.9	0.4	3.6%

5 ng/mL Cutoff		Within R	un (N=22)	Run-to-Run (N=88)	
Concentration	% of Cutoff	# Samples	EIA Result	# Samples	EIA Result
0 ng/mL	0 %	22	22 Neg	88	88 Neg
1.25 ng/mL	25 %	22	22 Neg	88	88 Neg
2.5 ng/mL	50 %	22	22 Neg	88	88 Neg
3.75 ng/mL	75 %	22	22 Neg	88	88 Neg
5 ng/mL	100 %	22	16 Neg/ 6 Pos	88	72 Neg/ 16 Pos
6.25 ng/mL	125 %	22	22 Pos	88	88 Pos
7.5 ng/mL	150 %	22	22 Pos	88	88 Pos
8.75 ng/mL	175 %	22	22 Pos	88	88 Pos
10 ng/mL	200 %	22	22 Pos	88	88 Pos

**Accuracy:** One hundred (100) unaltered clinical urine specimens were tested with the LZI Fentanyl II (Semi-Quantitative) Enzyme Immunoassay and confirmed by LC/MS. Specimens having a norfentanyl concentration equal to or greater than 5 ng/mL by LC/MS are defined as positive, and specimens with norfentanyl concentrations below 5 ng/mL by LC/MS are defined as negative in the table below. Near cutoff samples are defined as  $\pm 50$ % of the cutoff value. The correlation results are summarized as follows:

Semi-Quantitative Accuracy Study:

5 ng/mL Cutoff	Neg	< 50 % of the cutoff	Near Cutoff Neg	Near Cutoff Pos	High Pos	% Agree- ment
Positive	0	0	8**	10	40	100.0 %
Negative	20	20	2	0	0	84.0 %

The following table summarizes the result for the discordant samples:

5 ng/mL Cutoff	NFEN LC/MS (ng/mL)	LC/MS	LZI EIA (ng/mL)	LZI EIA
41**	2.7	-	6.6	+
43**	3.0	-	14.0	+
44**	3.0	-	6.5	+
45**	3.3	-	7.1	+
46**	3.5	1	10.2	+
47**	3.8	1	14.3	+
48**	3.9	-	6.9	+
49**	4.2	-	20.3	+

<sup>\*\*</sup> Discrepant between 50% below the cutoff concentration and cutoff (2.5 ng/mL - 4.9 ng/mL)

## Qualitative Accuracy Study:

5 ng/mL Cutoff	Neg	< 50 % of the cutoff		Near Cutoff Pos	High Pos	% Agree- ment
Positive	0	1*	8**	10	40	100.0 %
Negative	20	19	2	0	0	82.0 %

The following table summarizes the results for the qualitative discordant samples:

	Sample #	Norfentanyl LC/MS (ng/mL)	LC/MS Pos/Neg Result	AU480 EIA Qualitative Result (mAU)	AU480 EIA Qualitative Cutoff Rate (mAU)	LZI FEN II EIA Pos/Neg Result
Ī	37*	1.5	-	85.9	83.0	+
Ī	41**	2.7	-	111.3	83.0	+
	43**	3.0	-	207.9	83.0	+
	44**	3.0	-	107.7	83.0	+
Ī	45**	3.3	-	124.7	83.0	+
Ī	46**	3.5	-	169.6	83.0	+
Ī	47**	3.8	-	204.6	83.0	+
	48**	3.9	-	113.6	83.0	+
ſ	49**	4.2	-	263.1	83.0	+

<sup>\*</sup> Discrepant below 50% of the cutoff concentration (0 ng/mL - 2.49 ng/mL)

Discrepant samples contained levels of fentanyl that contributed to the false positive result.

<sup>\*\*</sup> Discrepant between 50% below the cutoff concentration and cutoff (2.5 ng/mL - 4.9 ng/mL)

**Specificity:** Various potentially interfering substances were tested for cross-reactivity with the assay. Test compounds were spiked into the drug-free urine calibrator matrix to various concentrations and evaluated against the cutoff calibrator

The following table lists the concentration of each test compound that gave a response approximately equivalent to that of the cutoff calibrator (as positive) or the maximal concentration of the compound tested that gave a response below the response of the cutoff calibrator (as negative). Compounds tested at high concentration with results below the cutoff value were listed as Not Detected (ND).

## Fentanyl and Metabolites:

Compound	Concentration Tested (ng/mL)	% Cross- Reactivity	Result
Fentanyl	3.8	131.58 %	Positive
Norfentanyl	5	100.00 %	Positive

#### **Structurally Related Compounds:**

Compound	Concentration Tested (ng/mL)	% Cross-Reactivity
4-Fluoro-Isobutyryl Fentanyl	20	25.00 %
9-Hydroxy Risperidone	100000	ND
Acetyl Fentanyl	7	71.43 %
Acetyl Norfentanyl	100	5.00 %
Acryl Fentanyl	4	125.00 %
Alfentanil	100000	ND
Butyryl Fentanyl	6	83.33 %
Butyryl Norfentanyl	40	12.50 %
Carfentanil Oxalate	100000	ND
Cis- d,I 3-Methyl Fentanyl	8	62.50 %
Cyclopropyl Fentanyl	3.2	156.25 %
Cyclopropyl Norfentanyl	25	20.00 %
Despropionyl Fentanyl (4-ANPP)	100000	ND
Furanyl Fentanyl	5.5	90.91 %
Furanyl Norfentanyl	180	2.78 %
(±) β-Hydroxy ThioFentanyl	5	100.00 %
Isobutyryl Fentanyl	15	33.33 %
Isobutyryl Norfentanyl	500	1.00 %
Labetalol Hydrochloride	100000	ND
Methoxyacetyl Fentanyl	3.5	142.86 %
MT-45	100000	ND
N-benzyl Furanyl Norfentanyl	11	45.45 %
N-benzyl Para-fluoro Norfentanyl	4	125.00 %
Norcarfentanil Oxalate	100000	ND
Ocfentanil	3.8	131.58 %
Para-fluoro Butyrl Fentanyl (p-FBF)	4.5	111.11 %
Para-fluoro Fentanyl	3.2	156.25 %
Remifentanil	100000	ND
Risperidone	100000	ND
Sufentanil	100000	ND
Thienyl Fentanyl	4	125.00 %
Thiofentanyl	3.2	156.25 %
Trans- d,I 3-Methyl Fentanyl	6	83.33 %
Trazodone	100000	ND
U-47700	100000	ND
Valeryl Fentanyl	70	7.14 %

# ${\bf Structurally\ Unrelated\ Compounds:}$

,	6 9 161	Spiked Norfentanyl Concentration			
Compound Spiked (ng/m		0 ng/mL	3.75 ng/mL Control	6.25 ng/mL Control	
Acetaminophen	100000	ND	Neg	Pos	
6-Acetylmorphine	100000	ND	Neg	Pos	
Acetylsalicylic Acid	100000	ND	Neg	Pos	
Amitriptyline	100000	ND	Neg	Pos	
Amlodipine Besylate	100000	ND	Neg	Pos	
Amoxicillin	100000	ND	Neg	Pos	
d-Amphetamine	100000	ND	Neg	Pos	
Atorvastatin	100000	ND	Neg	Pos	
Benzoylecgonine	100000	ND	Neg	Pos	
Buprenorphine	100000	ND	Neg	Pos	
Bupropion	100000	ND	Neg	Pos	
Caffeine	100000	ND	Neg	Pos	
Carbamazepine	100000	ND	Neg	Pos	
Cetirizine	100000	ND	Neg	Pos	
Chlorpheniramine	100000	ND	Neg	Pos	
Chlorpromazine	100000	ND	Neg	Pos	
Clomipramine	100000	ND	Neg	Pos	
Codeine	100000	ND	Neg	Pos	
Desipramine	100000	ND	Neg	Pos	
Dextromethorphan	40000	0.01 %	Pos	Pos	
Diphenhydramine	100000	ND	Neg	Pos	
Duloxetine	100000	ND	Neg	Pos	
Fluoxetine	100000	ND	Neg	Pos	
Fluphenazine	100000	ND	Neg	Pos	

## **Structurally Unrelated Compounds, continued:**

	Spiked [ ]	Spiked No	rfentanyl Cor	centration
Compound	(ng/mL)	0 ng/mL	3.75 ng/mL Control	6.25 ng/mL Control
Gabapentin	100000	ND	Neg	Pos
Hydrocodone	100000	ND	Neg	Pos
Hydromorphone	100000	ND	Neg	Pos
Ibuprofen	100000	ND	Neg	Pos
Imipramine	100000	ND	Neg	Pos
Lisinopril	100000	ND	Neg	Pos
Losartan	100000	ND	Neg	Pos
Loratadine	100000	ND	Neg	Pos
MDA (3,4-methylene- dioxyamphetamine)	100000	ND	Neg	Pos
MDEA	100000	ND	Neg	Pos
MDMA (3,4-methylene- dioxymethamphetamine)	100000	ND	Neg	Pos
Meperidine	100000	ND	Neg	Pos
Metformin	100000	ND	Neg	Pos
Metoprolol	100000	ND	Neg	Pos
Methadone	100000	ND	Neg	Pos
d-Methamphetamine	100000	ND	Neg	Pos
Morphine	100000	ND	Neg	Pos
Nalmefene	100000	ND	Neg	Pos
Nicotine	100000	ND	Neg	Pos
Nortriptyline	100000	ND	Neg	Pos
Omeprazole	100000	ND	Neg	Pos
Oxazepam	100000	ND	Neg	Pos
Oxycodone	100000	ND	Neg	Pos
Oxymorphone	100000	ND	Neg	Pos
Phenobarbital	100000	ND	Neg	Pos
(1S,2S)-(+)Pseudoephedrine	100000	ND	Neg	Pos
Quetiapine	100000	ND	Neg	Pos
Ranitidine	100000	ND	Neg	Pos
Salbutamol (Albuterol)	100000	ND	Neg	Pos
Sertraline	100000	ND	Neg	Pos
THC-COOH (11-Nor-Delta-9-THC-9- carboxylic acid)	100000	ND	Neg	Pos
l-Thyroxine	100000	ND	Neg	Pos
Tramadol	100000	ND	Neg	Pos
Zolpidem	100000	ND	Neg	Pos
Phencyclidine	100000	ND	Neg	Pos

It is possible that other substances and/or factors not listed above may interfere with the test and cause false positive results.

The following structurally unrelated compound which showed interference at  $\pm 25~\%$  of cutoff concentrations was then spiked into pooled negative human urine at  $\pm 50~\%$  of cutoff concentrations (2.5 ng/mL and 7.5 ng/mL) for the assay. Interference was still observed with dextromethorphan. Results are summarized in the following table:

Compound	Spiked [ ]	Spiked Norfentanyl Concentration		
Compound	(ng/mL)	0 ng/mL	2.5 ng/mL	7.5 ng/mL
Dextromethorphan	40000	Pos	Pos	Pos

## **Endogenous and Preservative Compound Interference Study:**

The following endogenous compounds were spiked into pooled negative human urine and the two levels of controls (3.75 ng/mL and 6.25 ng/mL) for the assay. The spiked solution was evaluated against cutoff calibrator. Interference was observed with Boric Acid. No other major interference with these compounds at physiological relevant concentrations as all spiked samples gave correct corresponding preliminary positive/negative results against the cutoff value of 5 ng/mL. Results are summarized in the following table:

	6 9 111	Spiked Norfentanyl Concentration			
Endogenous Substance	Spiked [ ] (mg/dL)	0 ng/mL	3.75 ng/mL Control	6.25 ng/mL Control	
Acetone	1000	Neg	Neg	Pos	
Ascorbic Acid	500	Neg	Neg	Pos	
Bilirubin	2	Neg	Neg	Pos	
Biotin	0.5	Neg	Neg	Pos	
Boric Acid	1000	Neg	Neg	Neg	
Calcium Chloride (CaCl2)	300	Neg	Neg	Pos	
Citric Acid (pH 3)	200	Neg	Neg	Pos	
Creatinine	500	Neg	Neg	Pos	
Ethanol	1000	Neg	Neg	Pos	
Galactose	10	Neg	Neg	Pos	
γ-Globulin	500	Neg	Neg	Pos	
Glucose	3000	Neg	Neg	Pos	
Hemoglobin	300	Neg	Neg	Pos	
Human Serum Albumin	500	Neg	Neg	Pos	
Human Urine (pooled)	N/A	Neg	Neg	Pos	
β-hydroxybutyric Acid	100	Neg	Neg	Pos	
Oxalic Acid	100	Neg	Neg	Pos	
Potassium Chloride	1000	Neg	Neg	Pos	

#### **Endogenous and Preservative Compound Interference Study, continued:**

	C93 []	Spiked Norfentanyl Concentration			
Endogenous Substance	Spiked [ ] (mg/dL)	0 ng/mL	3.75 ng/mL Control	6.25 ng/mL Control	
Riboflavin	7.5	Neg	Neg	Pos	
Sodium Azide	1000	Neg	Neg	Pos	
Sodium Chloride	1000	Neg	Neg	Pos	
Sodium Fluoride	1000	Neg	Neg	Pos	
Sodium Phosphate	300	Neg	Neg	Pos	
Urea	6000	Neg	Neg	Pos	
Uric Acid	10	Neg	Neg	Pos	
LZI Urine-Based Calibrator Buffer	N/A	Neg	Neg	Pos	

The following endogenous compounds which showed interference at  $\pm 25~\%$  of htet cutoff concentration were then spiked into negative urine and at  $\pm 50~\%$  of cutoff concentrations (2.5 ng/mL and 7.5 ng/mL) for the assay. Interference was still observed with Boric Acid at 1 % w/v. Results are summarized in the following table:

Endogonous Substance	Spiked []	Spiked Norfentanyl Concentration		
Endogenous Substance	(mg/dL)	0 ng/mL	2.5 ng/mL	7.5 ng/mL
Boric Acid	1000	Neg	Neg	Neg

**pH Interference Study:** Negative urine and urine spiked with analyte to the two levels of controls (3.75 ng/mL and 6.25 ng/mL) were adjusted to the following pH levels and tested by the assay. The pH adjusted solutions were evaluated against the cutoff calibrator.

No major interference with these pH levels was observed as all pH adjusted levels gave correct corresponding preliminary positive/negative results against the cutoff value of 5 ng/mL. Results are summarized in the following table:

	Spiked	Spiked Norfentanyl Concentration				
pН	0 ng/mL	3.75 ng/mL Control	6.25 ng/mL Control			
pH 3	Neg	Neg	Pos			
pH 4	Neg	Neg	Pos			
pH 5	Neg	Neg	Pos			
pH 6	Neg	Neg	Pos			
pH 7	Neg	Neg	Pos			
pH 8	Neg	Neg	Pos			
pH 9	Neg	Neg	Pos			
pH 10	Neg	Neg	Pos			
pH 11	Neg	Neg	Pos			

**Specific Gravity:** Samples ranging in specific gravity from 1.000 to 1.027 were split into three portions each and either left un-spiked or spiked to a norfentanyl concentration of either 3.75 or 6.25 ng/mL (the negative and positive control concentrations, respectively). These samples were then evaluated in qualitative mode. No interference was observed.

Specific	Spiked Norfentanyl Concentration				
Gravity	0 ng/mL	3.75 ng/mL Control	6.25 ng/mL Control		
1.000	Neg	Neg	Pos		
1.003	Neg	Neg	Pos		
1.005	Neg	Neg	Pos		
1.008	Neg	Neg	Pos		
1.010	Neg	Neg	Pos		
1.012	Neg	Neg	Pos		
1.015	Neg	Neg	Pos		
1.018	Neg	Neg	Pos		
1.020	Neg	Neg	Pos		
1.022	Neg	Neg	Pos		
1.025	Neg	Neg	Pos		
1.027	Neg	Neg	Pos		

Open-Vial Reagent and Calibrator/Control Stability: Real-time data for open-vial reagent and calibrator/control stability studies at Cold Temperature (2-8°C) have been carried out up to Day 730. Results from open-vial studies indicate that degradation is minimal up to Day 730, and, based on the real-time data, suggests an open-vial stability of up to 24 months. Open-vial reagents and calibrators/controls should be stored at 2-8°C for maximum shelf life

Closed-Vial Calibrator/Control Stability: Real-time data for closed-vial calibrator/control stability studies at Cold Temperature (2-8°C) have been carried out up to Day 730. Results from closed-vial studies indicate that degradation is minimal at Cold Temperature (2-8°C) up to Day 730 in comparison to Day 1. Closed-vial calibrators/controls should be stored at 2-8°C for maximum shelf life.

## Symbols Used

EC REP	Authorized Representative	REAGENT 1	R <sub>1</sub> , Antibody/ Substrate Reagent
8	Biological Risks	REAGENT 2	R <sub>2</sub> , Enzyme- Drug Conjugate Reagent
C€	CE Mark	REF	Reference Number
[]i	Consult Instructions for Use	SDS	Safety Data Sheet
CONTENTS	Contents	2°C √8°C	Temperature Limits
GTIN	Global Trade Item Number	T.K.	Test Kit Number
LOT	Lot Number	Ξ	Use-by Date
444	Manufacturer		

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A point (period/stop) is always used in this instruction for use document as the decimal separator to mark the border between the integral and the fractional parts of a decimal numeral. Separators for thousands are not used.

Additions, deletions, or changes are indicated by a change bar in the margin. For technical assistance please call: (408) 970-8811

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