# LZI Cotinine Enzyme Immunoassay

**REF** 0230 (100/37.5 mL R<sub>1</sub>/R<sub>2</sub> Kit) 0231 (1000/375 mL R<sub>1</sub>/R<sub>2</sub> Kit)

# Lin-Zhi International, Inc.

### **Intended Use**

The Lin-Zhi International, Inc. (LZI) Cotinine Enzyme Immunoassay is intended for the qualitative and semi-quantitative determination of cotinine in human

 urine at a cutoff value of 500 ng/mL. The assay is designed for use with a number of automated clinical chemistry analyzers. This is a Non-FDA Approved
assay for Employment and Insurance Testing Use Only and as such should not
be repackaged for in vitro diagnostic use.

 The assay provides only a preliminary analytical result. A more specific alternative analytical chemistry method must be used in order to obtain a

confirmed analytical result. Gas or Liquid Chromatography/Mass Spectrometry (GC/MS or LC/MS) are the preferred confirmatory methods (1, 2). Clinical consideration and professional judgment should be exercised with any immunoassay result, particularly when the preliminary test result is positive.

### **Summary and Explanation of Test**

Nicotine is the primarily addictive compound in tobacco products (3). For the past three decades, there has been a tremendous attention to tobacco smoking and the so-called "passive inhalation" of tobacco smoke due to its correlation to lung cancer.

Inhaled tobacco smoke reaches small airways and alveoli of the lungs, where 90 % of nicotine is absorbed. When nicotine is absorbed, it is readily metabolized into cotinine by the liver (4). Urine concentrations of both nicotine and cotinine correlate with cigarette use in active smokers (5). While nicotine has a very short half-life of approximately 40 minutes (6), cotinine has an average half-life of 20 hours (7), and can be detected in the urine of a smoker even several days after the smoking has ceased.

Several methods have been used to determine the smoking status of an individual. These include measurement of thiocyanate, carbon monoxide, and cotinine. Measurement of both thiocyanate and carbon monoxide, however, is more likely to be affected by environmental factors and can cause false positive results. Since cotinine can only be derived from metabolism of nicotine, it is a better marker for determination of the smoker status.

### **Assay Principle**

The LZI Cotinine assay is a homogeneous enzyme immunoassay ready-to-use liquid reagent. The assay is based on competition between drug in the sample and drug labeled with the enzyme glucose-6-phosphate dehydrogenase (G6PDH) for a fixed amount of antibody in the reagent (8). Enzyme activity decreases upon binding to the antibody, and the drug concentration in the sample is measured in terms of enzyme activity.

In the absence of drug in the sample, cotinine-labeled G6PDH conjugate is bound to antibody, and the enzyme activity is inhibited. On the other hand, when drug is present in the sample, antibody binds to the free drug; the unbound cotinine-labeled G6PDH then exhibits its maximal enzyme activity. Active enzyme converts nicotinamide adenine dinucleotide (NAD) to NADH, resulting in an absorbance change that can be measured spectrophotometrically at a 340 nm primary wavelength.

### **Reagents Provided**

Antibody/Substrate Reagent ( $R_1$ ): Contains mouse monoclonal anti-cotinine antibody, glucose-6-phosphate (G6P), nicotinamide adenine dinucleotide (NAD), stabilizers, and sodium azide (0.09 %) as a preservative. Enzyme-drug Conjugate Reagent ( $R_2$ ): Contains glucose-6-phosphate dehydrogenase (G6PDH) labeled with cotinine in buffer with sodium azide (0.09 %) as a preservative.

# **Calibrators and Controls\***

\*Calibrators and controls are sold separately and contain negative human urine with sodium azide as a preservative.

COTININE Calibrators/Controls	REF
Negative Calibrator	0001
Low Calibrator/Level 1 Control: 250 ng/mL cotinine	0232
Cutoff Calibrator: 500 ng/mL cotinine	0233
Intermediate Calibrator: 1000 ng/mL cotinine	0234
High Calibrator: 2000 ng/mL cotinine	0235
Level 2 Control: Contains 750 ng/mL cotinine	0238

### **Precautions and Warning**

• This test is for Employment and Insurance Testing Use Only. This test should not be re-packaged for in vitro diagnostic use.

For Employment and Insurance Testing Use Only

- Harmful if swallowed.
- Reagent contains sodium azide as a preservative, which may form explosive compounds in metal drain lines. When disposing such reagents or wastes always flush with a large volume of water to prevent azide build-up. See National Institute for Occupational Safety and Health Bulletin: Explosive Azide Hazards (9).
- Do not use the reagents beyond their expiration dates.

### **Reagent Preparation and Storage**

The reagents are ready-to-use. No reagent preparation is required. All assay components should be refrigerated at 2-8°C when not in use.

### Specimen Collection and Handling

Urine samples may be collected in plastic or glass containers. Some plastics

- | may absorb drugs. Use of plastics such as polyethylene is recommended (10). Use fresh urine specimens for the test. If a sample cannot be analyzed
- immediately, it may be refrigerated at 2-8°C for up to seven days (11, 12). For longer storage, keep sample frozen at -20°C and then thaw before use. Studies have shown cotinine analytes in urine are stable at -20°C for up to seven
- weeks (12, 13). Samples should be at room temperature (18-25°C) for testing. Samples with high turbidity should be centrifuged before analysis. Adulteration may cause erroneous results. If sample adulteration is suspected, obtain a new sample and forward both samples to the laboratory for testing. *Handle all urine specimens as if they are potentially infectious.*

### Instrument

Clinical chemistry analyzers capable of maintaining a constant temperature, pipetting samples, mixing reagents, measuring enzyme rates at a 340 nm primary wavelength and timing the reaction accurately can be used to perform this homogeneous immunoassay.

Performance characteristics presented in this package insert have been validated on the Hitachi 717. If other instruments are used, performance will need to be validated by the laboratory (14, 15).

# Assay Procedure

Analyzers with the specifications indicated above are suitable for performing this homogeneous enzyme immunoassay. Refer to the specific parameters used for each analyzer before performing the assay. Typical assay parameters used for the Hitachi 717 analyzer include a 20  $\mu$ L sample, 200  $\mu$ L of antibody reagent (R<sub>1</sub>), and 75  $\mu$ L of enzyme conjugate reagent (R<sub>2</sub>) at 37°C incubation temperature, 26-30 reading frames, and a 340 nm primary wavelength. For qualitative analysis, use 500 ng/mL as the cutoff calibrator. For semi-quantitative analysis, use all five calibrators. Recalibration should be performed after reagent bottle change or if there is a change in calibrators or reagent lot. Two levels of controls are also available for monitoring the reference point: use the 250 ng/mL and 750 ng/mL for the 500 ng/mL cutoff level.

### **Calibration and Quality Control**

Good laboratory practices recommend the use of at least two levels of control specimens (one positive and one negative control near the cutoff) to ensure proper assay performance. Controls should be run with each new calibration and after specific maintenance or troubleshooting procedures as detailed in the instrument system manual. Each laboratory should establish its own control frequency. If any trends or sudden change in control value are observed, review all operating parameters, or contact LZI technical support for further assistance. Laboratories should comply with all federal, state, and local laws, as well as guidelines and regulations.

#### Results

| Note: A preliminary positive test result does not necessarily mean a person is a smoker and a negative test result does not necessarily mean a person did not smoke. There are a number of factors that influence the reliability of immunoassays.



**Qualitative:** The urine concentration of cotinine for non-smokers is normally well below 500 ng/mL (4, 5). The urine concentration of cotinine for active smoker averages from 1000 ng/mL to as high as 8000 ng/mL (5). A sample with a change in absorbance ( $\Delta$ mA/min) equal to or greater than that obtained

with the cutoff calibrator is considered a preliminary positive. A sample with a change in absorbance ( $\Delta mA/min$ ) lower than that obtained with the calibrator used as the cutoff is considered negative.

**Semi-Quantitative**: The semi-quantitative mode is for purposes of (1) enabling laboratories to determine an appropriate dilution of the specimen for verification by a confirmatory method such as GC/MS, LC/MS or (2) permitting laboratories to establish quality control procedures. When an approximation of concentration is required, a calibration curve can be established with five calibrators. The concentration of cotinine in the sample may then be estimated from the calibration curve.

### Limitations

- 1. A preliminary positive result from the assay indicates only the presence of cotinine. The test is not intended for quantifying this single analyte in samples.
- 2. A preliminary positive result does not necessarily indicate a person is a smoker.
- 3. A negative result does not necessarily mean a person did not smoke.
- 4. Care should be taken when reporting results as numerous factors (e.g., fluid intake, endogenous or exogenous interferents) may influence the urine test result.
- 5. Preliminary positive results should be confirmed by other affirmative, analytical chemistry methods (e.g., chromatography), preferably GC/MS or LC/MS.
- 6. The test is designed for use with human urine only.
- 7. The test is not for therapeutic drug monitoring.

### **Typical Performance Characteristics**

The results shown below were performed with a single Hitachi 717 automated clinical chemistry analyzer.

#### Precision:

<u>Qualitative analysis</u>: The three calibrators and two levels of controls were evaluated. Typical results ( $\Delta$ mA/min) are as follows:

Concentration	Wit	hin Run (N	=21)	Run-to-Run* (N=12)		
Concentration	Mean	SD	% CV	Mean	SD	% CV
Negative	414.6	2.2	0.5 %	412.1	2.9	0.7 %
250 ng/mL	456.6	3.5	0.8 %	453.1	4.1	0.9 %
500 ng/mL	483.6	3.2	0.7 %	483.4	2.6	0.5 %
750 ng/mL	508.8	2.8	0.6 %	507.5	2.1	0.4 %
2000 ng/mL	593.4	3.9	0.7 %	589.1	5.4	0.9 %

\* Run-to-Run testing completed over 2 weeks.

<u>Semi-quantitative analysis</u>: The concentrations of 500 ng/mL and the two levels of controls were determined with reference curve from five calibrators. Typical results (ng/mL) are as follows:

Concentration	Within Run (N=21)			Run-to-Run* (N=12)		
Concentration	Mean	SD	% CV	Mean	SD	% CV
250 ng/mL	252.8	9.5	3.8 %	256.0	14.9	5.8 %
500 ng/mL	505.7	13.3	2.6 %	503.9	27.8	5.5 %
750 ng/mL	755.4	17.1	2.3 %	742.5	17.6	2.4 %
* Pup to Pup tasting completed over 2 weeks						

\* Run-to-Run testing completed over 2 weeks.

**Sensitivity:** Sensitivity, defined as the lowest concentration that can be differentiated from negative urine with 95 % confidence, was tested to be 25 ng/mL.

**Specificity:** Various potentially interfering substances were tested for crossreactivity with the assay. Test compounds were spiked into the drug-free urine calibrator matrix to various concentrations and evaluated against the cutoff calibrator.

The following table lists the concentration of each test compound that gave a response approximately equivalent to that of the cutoff calibrator (as positive) or the maximal concentration of the compound tested that gave a response below the response of the cutoff calibrator (as negative).

#### **Structurally Related Cotinine Compounds:**

Compound	Target [] (ng/mL)	Observed Value (ng/mL)	% Cross- Reactivity
(R,S)-Norcotinine	62,500	571.7	0.91 %
S(-)-Nicotine	250,000	204.2	0.08 %
(+)-Anabasine	250,000	22.3	0.01 %
(+/-)-Nornicotine	250,000	35.8	0.01 %
Trans-3'-Hydroxycotinine	15,625	853.5	5.46 %

#### Structurally Unrelated Compounds:

	Test	% Cross-Reactivity Spiked Cotinine Concentration			
	[]	Spi			
Compound		0	(-25% Cutoff)	(+25% Cutoff)	
	(ng/mL)	ng/mL	375 ng/mL	625 ng/mL	
A	25.000	0	Control	Control	
Acetaminophen	35,000	Neg	Neg	Pos	
6-Acetylmorphine	10,000	Neg	Neg	Pos	
Acetylsalicylic Acid	35,000	Neg	Neg	Pos	
Albuterol (Salbutamol)	35,000	Neg	Neg	Pos	
Amitriptyline	35,000	Neg	Neg	Pos	
d-Amphetamine	100,000	Neg	Neg	Pos	
Benzoylecgonine	35,000	Neg	Neg	Pos	
Buprenorphine	15,000	Neg	Neg	Pos	
Bupropion	35,000	Neg	Neg	Pos	
Caffeine	35,000	Neg	Neg	Pos	
Carbamazepine	50,000	Neg	Neg	Pos	
Cetirizine	35,000	Neg	Neg	Pos	
Chlorpheniramine	35,000	Neg	Neg	Pos	
Chlorpromazine	35,000	Neg	Neg	Pos	
Clomipramine	100,000	Neg	Neg	Pos	
Codeine	100,000	Neg	Neg	Pos	
Desipramine	35,000	Neg	Neg	Pos	
Diphenhydramine	100,000	Neg	Neg	Pos	
Ephedrine	100,000	Neg	Neg	Pos	
Fentanyl	10,000	Neg	Neg	Pos	
Fluoxetine	35,000	Neg	Neg	Pos	
Fluphenazine	50,000	Neg	Neg	Pos	
Hydrocodone	35,000	Neg	Neg	Pos	
Hydromorphone	50,000	Neg	Neg	Pos	
Ibuprofen	35,000	Neg	Neg	Pos	
Imipramine	35,000	Neg	Neg	Pos	
Lidocaine	35,000	Neg	Neg	Pos	
Loratadine		Neg		Pos	
	50,000	0	Neg		
Maprotiline	50,000	Neg	Neg	Pos	
MDA (3,4-	100,000	Neg	Neg	Pos	
methylenedioxyamphetamine)	100.000	-	-	D	
MDEA	100,000	Neg	Neg	Pos	
MDMA (3,4-	100,000	Neg	Neg	Pos	
methylenedioxyamphetamine)	100.000	-	-		
Meperidine	100,000	Neg	Neg	Pos	
Methadone	35,000	Neg	Neg	Pos	
d-Methamphetamine	100,000	Neg	Neg	Pos	
Methapyrilene	35,000	Neg	Neg	Pos	
Methaqualone	35,000	Neg	Neg	Pos	
Metronidazole	35,000	Neg	Neg	Pos	
Morphine	35,000	Neg	Neg	Pos	
Nicotine	100,000	Neg	Neg	Pos	
Nortriptyline	35,000	Neg	Neg	Pos	
Oxazepam	100,000	Neg	Neg	Pos	
Oxycodone	35,000	Neg	Neg	Pos	
Oxymorphone	100,000	Neg	Neg	Pos	
Phencyclidine	10,000	Neg	Neg	Pos	
Pentazocine	20,000	Neg	Neg	Pos	
Phenobarbital	100,000	Neg	Neg	Pos	
d-Propoxyphene					
(Dextropropoxyphene)	35,000	Neg	Neg	Pos	
Propranolol	35,000	Neg	Neg	Pos	
Ranitidine	50,000	Neg	Neg	Pos	
Sertraline	35,000	Neg	Neg	Pos	
THC-COOH	22,000			- 00	
(11-Nor-Delta-9-THC-9-	1000	Neg	Neg	Pos	
Carboxylic Acid)	1000	1.05	1.05	100	
Thioridazine	50,000	Neg	Neg	Pos	
Tramadol	50,000	Neg	Pos	Pos	
	50,000	Neg	Neg	Pos	

It is possible that other substances and/or factors not listed above may interfere with the test and cause false positive results.

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Additions, deletions, or changes are indicated by a change bar in the margin. For technical assistance please call: (408) 970-8811

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